

Rapport remis à titre d'exemple, valide en date du 14/03/2022

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Lung adenocarcinoma
NAME
DATE OF BIRTH
SEX
MEDICAL RECORD #

ORDE MEDIO

ORDERING PHYSICIAN
MEDICAL FACILITY
ADDITIONAL RECIPIENT
MEDICAL FACILITY ID
PATHOLOGIST

SI SI D

SPECIMEN SITE
SPECIMEN ID
SPECIMEN TYPE
DATE OF COLLECTION
SPECIMEN RECEIVED

Genomic Signatures

Microsatellite status - MS-Stable
Tumor Mutational Burden - 8 Muts/Mb

Gene Alterations

For a complete list of the genes assayed, please refer to the Appendix.

EGFR exon 19 deletion (T751_I759>N)

CCND1 amplification

CDK4 amplification - equivocal

FGF19 amplification - equivocal[†]

FGF3 amplification - equivocal

FGF4 amplification - equivocal

KDM6A rearrangement intron 10, rearrangement exon

23

NFKBIA amplification - equivocal

NKX2-1 amplification - equivocal

TP53 C176Y

7 Disease relevant genes with no reportable alterations: ALK, BRAF, ERBB2, KRAS, MET, RET, ROS1

† See About the Test in appendix for details.

Report Highlights

- Targeted therapies with NCCN categories of evidence in this tumor type: Afatinib (p. 9), Dacomitinib (p. 10), Erlotinib (p. 10), Gefitinib (p. 11), Osimertinib (p. 12)
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 13)

GENOMIC SIGNATURES

Microsatellite status - MS-Stable

Tumor Mutational Burden - 8 Muts/Mb

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Genomic Signatures section

No therapies or clinical trials. see Genomic Signatures section

(i)	FOUNDATIONONE®CDX

GENE ALTERATIONS	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)		
EGFR - exon 19 deletion (T751_I759>N)	Afatinib 1	none		
	Dacomitinib 1			
	Erlotinib 1			
	Gefitinib 1			
10 Trials see <i>p.</i> 17	Osimertinib 1			
CCND1 - amplification	none	none		
9 Trials see p. 13				
CDK4 - amplification - equivocal	none	none		
10 Trials see p. 15				
		NCCN category		

GENE ALTERATIONS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Alterations section.

FGF19 - amplification - equivocal	p. 5	rearrangement exon 23	p. 6
FGF3 - amplification - equivocal	p. 6	NFKBIA - amplification - equivocal	p. 7
FGF4 - amplification - equivocal	p. 6	NKX2-1 - amplification - equivocal	p. 7
KDM6A - rearrangement intron 10,		TP53 - C176Y	p. 8

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved through a centralized EU procedure or a national procedure in an EU Member State. Therapies, including but not limited to the following, have been approved nationally and may not be available in all EU Member States: Tretinoin, Anastrozole, Bicalutamide, Cyproterone, Exemestane, Flutamide, Goserelin, Letrozole, Leuprorelin, Triptorelin.



GENOMIC SIGNATURES

GENOMIC SIGNATURE

Microsatellite status

RESULT MS-Stable

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, p=0.001)⁵.

FREQUENCY & PROGNOSIS

MSI-H is generally infrequent in NSCLC, reported in fewer than 1% of samples across several large studies⁶⁻¹¹, whereas data on the reported incidence of MSI-H in SCLC has been limited and conflicting¹²⁻¹⁵. One study reported MSI-H in lung adenocarcinoma patients with smoking history, and 3 of 4 MSI-H patients examined also had metachronous carcinomas in other organs, although this has not been investigated in large scale studies⁶. Published data investigating the prognostic implications of MSI in NSCLC are limited (PubMed, Oct 2021).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁶. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH₂, MSH₆, or PMS₂¹⁶⁻¹⁸. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers19-21. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{16,18,20-21}.

GENOMIC SIGNATURE

Tumor Mutational Burden

RESULT 8 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²²⁻²⁴, anti-PD-1 therapies²²⁻²⁵, and combination nivolumab and ipilimumab²⁶⁻³¹. Multiple clinical trials of PD-1- or PD-L1-targeting immune checkpoint inhibitors or combination of PD-1 and CTLA-4 inhibitors in NSCLC have reported that patients with tumors harboring TMB ≥10 Muts/Mb derive greater clinical benefit from these therapies than those with TMB <10 Muts/Mb (based on this assay or others); similarly, higher efficacy of anti-PD-1 or anti-PD-L1 immunotherapy for treatment of patients with NSCLC, compared with the use of chemotherapy, has been observed more significantly in cases of TMB ≥10 Muts/Mb (based on this assay or others);^{22-23,26-28,32-39}. Improved OS of patients with

NSCLC treated with pembrolizumab plus chemotherapy relative to chemotherapy only⁴⁰, or those treated with nivolumab plus ipilimumab also relative to chemotherapy⁴¹, has been observed across all TMB levels.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that unspecified lung non-small cell lung carcinoma (NSCLC), lung adenocarcinoma, and lung squamous cell carcinoma (SCC) samples harbored median TMBs between 6.3 and 9 Muts/Mb, and 12% to 17% of cases had an elevated TMB of greater than 20 Muts/Mb⁴². Lower TMB is observed more commonly in NSCLCs harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are commonly observed in elevated TMB cases⁴³. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC44-45, several other large studies did find a strong association with increased TMB⁴⁶⁻⁴⁹. TMB >10 muts/Mb was found to be more frequent in NSCLC metastases compared with primary tumors for both adenocarcinoma (38% vs. 25%) and SCC (41% vs. 35%) subtypes⁵⁰. A large study of Chinese patients with lung adenocarcinoma reported a shorter median OS for tumors with a higher number of mutations in a limited gene set compared with a

lower mutation number (48.4 vs. 61.0 months)⁴⁴. Another study of patients with NSCLC correlated elevated TMB with poorer prognosis and significantly associated lower TMB in combination with PD-L1 negative status with longer median survival in patients with lung adenocarcinoma⁵¹. However, no significant prognostic association of TMB and/or PD-L1 status with survival has been reported in patients with lung SCC⁵¹⁻⁵².

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁵³⁻⁵⁴ and cigarette smoke in lung cancer^{32,55}, treatment with temozolomide-based chemotherapy in glioma⁵⁶⁻⁵⁷, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁵⁸⁻⁶², and microsatellite instability (MSI)^{58,61-62}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{22-23,26-28,32-39,63}.

GENOMIC FINDINGS

GENE

EGFR

ALTERATION exon 19 deletion (T751_I759>N)

TRANSCRIPT ID NM_005228

CODING SEQUENCE EFFECT 2252 2276>A

VARIANT ALLELE FREQUENCY (% VAF) 24.6%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

For patients with non-small cell lung cancer, EGFR activating mutations may predict sensitivity to EGFR TKIs, including erlotinib⁶⁴, gefitinib⁶⁵, afatinib⁶⁶, dacomitinib⁶⁷, and osimertinib⁶⁸; however, the data for patients with other tumor types are limited⁶⁹⁻⁷⁴. The Phase 1 CHRYSALIS study of amivantamab monotherapy or in combination with lazertinib for the treatment of EGFR-mutated non-small cell lung cancer (NSCLC) has produced encouraging preliminary results for treatment-naive patients and patients who relapsed after treatment with osimertinib with and without chemotherapy, including

osimertinib-relapsed patients with biomarkers indicating EGFR/MET-based osimertinib resistance75-78. In a Phase 1 trial, the HER3-targeted antibody patritumab deruxtecan elicited an ORR of 39% (22/57, 1 CR) and a median PFS of 8.2 months for patients with non-small cell lung cancer previously treated with an EGFR TKI, many of whom displayed TKI resistance alterations⁷⁹. A Phase 1 trial evaluating the EGFR inhibitor AZD3759 reported a reduction in the volume of brain metastases in 40% (8/20) of patients with previously treated non-small cell lung cancer (NSCLC) harboring either the EGFR L858R alteration or EGFR exon 19 deletion, including 3 confirmed PRs and 3 unconfirmed PRs80-81. In a Phase 1/2 trial for advanced NSCLC, the brain-penetrant third-generation EGFR TKI lazertinib enabled ORRs of 54% (69/127) for all evaluable patients and 44% (8/18, intracranial) for patients with brain metastases⁸². The Phase 3 IMpower150 study showed that the addition of atezolizumab to bevacizumab plus chemotherapy treatment also had clinical efficacy for patients with EGFR-mutated or ALK-rearranged metastatic NSCLC83; therefore, the patient's clinical context should be considered.

FREQUENCY & PROGNOSIS

EGFR mutation has been reported in 12-36% of

lung adenocarcinomas^{48,84-85} and in 4% of lung squamous cell carcinomas⁸⁶. EGFR protein expression/overexpression has been reported in up to 70% of NSCLC cases⁸⁷⁻⁹². In addition, expression of EGFR protein has been shown to be higher in lung squamous cell carcinoma samples as compared to lung adenocarcinoma⁹³⁻⁹⁴. In patients with lung adenocarcinoma, EGFR mutation was a predictor of poor overall survival⁹⁵⁻⁹⁶. However, EGFR mutations have been reported to predict improved survival in patients with resected Stage 1-3 lung adenocarcinoma⁹⁷ or resected Stage 1 NSCLC⁹⁸.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide⁹⁹. EGFR exon 19 deletion mutations, such as seen here, have been shown to activate the tyrosine kinase activity of EGFR and to confer sensitivity to EGFR tyrosine kinase inhibitors such as erlotinib, gefitinib¹⁰⁰⁻¹⁰², afatinib¹⁰³, osimertinib¹⁰⁴, and dacomitinib^{67,105}, although limited preclinical data suggest reduced sensitivity to lapatinib¹⁰⁶⁻¹⁰⁷.

CCND1

ALTERATION amplification

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies

Amplification or overexpression of CCND1 may predict sensitivity to CDK4/6 inhibitors, such as abemaciclib, palbociclib, and ribociclib¹⁰⁸⁻¹¹³, although as monotherapy these agents have shown limited activity in tumor types other than breast cancer^{112,114}. In refractory advanced solid tumors with CCND1 (n=39) or CCND3 (n=1) amplification and retinoblastoma protein expression, palbociclib resulted in SD for 39% (14/36) of patients and a median PFS of 1.8 months in

the NCI-MATCH trial¹¹⁵; 4 patients (13%, 4/36 overall) with squamous cell carcinomas (lung, esophageal, or laryngeal) or adenoid cystic carcinoma experienced prolonged SD in this study¹¹⁵. Among 9 patients with CCND1-amplified advanced solid tumors, 1 patient with bladder cancer responded to ribociclib in a Phase 2 trial¹¹⁶.

Potential Resistance —

CCND1 amplification may predict worse outcomes on immune checkpoint inhibitors (anti-PD-1/PD-L1/CTLA-4) in solid tumors on the basis of 2 meta-analyses¹¹⁷⁻¹¹⁸; in these studies, CCND1 amplification was associated with significantly decreased response rate¹¹⁸ and OS (HR=1.6-2.0)¹¹⁷⁻¹¹⁸ across various tumor types and significantly shorter OS specifically in urothelial carcinoma (HR=2.2-3.6), melanoma (HR=1.6-2.5), and solid tumors harboring elevated TMB (HR=2.8)¹¹⁷⁻¹¹⁸.

FREQUENCY & PROGNOSIS

CCND1 amplification has been reported in 2-25% of lung adenocarcinoma^{48,85,119-120} and 6-38% of lung squamous cell carcinoma^{86,119-120} cases. Expression of cyclin D1 has been reported in 59% (36/61) of non-small cell lung cancer (NSCLC) tumors analyzed¹²¹. The prognostic significance of CCND1 amplification in NSCLC is not clear¹²². Cyclin D1 protein expression was not associated with clinicopathologic parameters of NSCLC in one study¹²¹.

FINDING SUMMARY

CCND1 encodes cyclin D1, a binding partner of the kinases CDK4 and CDK6, that regulates RB activity and cell cycle progression. Amplification of CCND1 has been positively correlated with cyclin D1 overexpression¹²³ and may lead to excessive proliferation¹²⁴⁻¹²⁵.

GENOMIC FINDINGS

CDK4

ALTERATION amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

CDK4 amplification or activation may predict sensitivity to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib^{109,111-112,126}. Clinical benefit has been reported for limited tumor types including patients with CDK4-amplified liposarcoma and sarcoma in response to treatment with abemaciclib¹²⁷,

palbociclib^{126,128}, and ribociclib¹¹⁶.

FREQUENCY & PROGNOSIS

In the TCGA datasets, CDK4 amplification or mutation occurs in 7% and 1% of lung adenocarcinoma cases, respectively¹²⁹; however, neither were detected in any of the lung squamous cell carcinoma cases⁸⁶. CDK4 amplification correlated with high CDK4 gene and protein expression in lung tumors¹³⁰. High CDK4 protein expression has been detected in 23-47% of nonsmall cell lung cancers, specifically in 38% (18/47) of lung adenocarcinomas, 44% (4/9) of lung squamous cell carcinomas, and 83% (10/12) of large cell lung cancers¹³⁰⁻¹³². A preclinical study suggests targeting of CDK4 as a potential strategy against KRAS-driven lung adenocarcinomas¹³³.

High CDK4 protein expression predicted poor overall survival in patients with lung cancer in one study¹³².

FINDING SUMMARY

CDK4 encodes the cyclin-dependent kinase 4, which regulates the cell cycle, senescence, and apoptosis¹³⁴. CDK4 and its functional homolog CDK6 are activated by D-type cyclins and promote cell cycle progression by inactivating the tumor suppressor Rb¹³⁵⁻¹³⁶. Amplification of the chromosomal region that includes CDK4 has been reported in multiple cancer types, including lung cancer, glioblastoma, and liposarcoma, and has been associated with overexpression of CDK4 protein^{126,130,137-142}.

GENE

FGF19

ALTERATION amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies that directly address genomic alterations in FGF19. However, FGF19 amplification predicts sensitivity to FGFR4 inhibitors in liver cancer cell lines143-144; selective FGFR4 inhibition reduced tumor burden in an FGF19-amplified HCC xenograft model¹⁴⁵. A Phase 1 study of the FGFR4 inhibitor fisogatinib (BLU-554) for patients with previously treated hepatocellular carcinoma (HCC), most of whom had received prior sorafenib treatment, reported a 16.7% ORR (11/66, 1 CR, ongoing for >1.5 years) and a median PFS of 3.3 months for FGF19-IHCpositive patients; poorer outcomes (o% ORR, PFS of 2.3 months) were observed for patients with negative or unknown FGF19 IHC scores146. Acquisition of FGFR4 mutations may represent a

mechanism of resistance for patients with FGF19 overexpression who initially responded but then progressed on fisogatinib¹⁴⁴. Preliminary results from the dose escalation part of a Phase 1/2 study evaluating another FGFR4 inhibitor, FGF401, showed an ORR of 7.6% (4/53), SD rate of 52.8% (28/53), and a median time to progression of 4.1 months; responses were observed in both FGF19-positive and FGF19-negative cases¹⁴⁷. In a retrospective analysis, a trend toward response to sorafenib treatment and FGF19 copy number gain was observed in patients with HCC, and 2 patients harboring FGF19 copy number gain experienced a CR148. A case study reported activity of pan-FGFR inhibitors in FGF-amplified cancers; following treatment with a selective pan-FGFR inhibitor, a patient with head and neck squamous cell carcinoma (HNSCC) and amplification of 11q13 (FGF3, FGF4, FGF19) and 12p13 (FGF6 and FGF23) experienced a radiologic CR149. Other therapies targeting FGF19 or FGFR4 signaling are in development150.

FREQUENCY & PROGNOSIS

In the TCGA datasets, FGF19 amplification has been reported with highest incidence in

esophageal carcinoma (34%), head and neck squamous cell carcinoma (23%), breast carcinoma (15%), lung squamous cell carcinoma (13%), and cholangiocarcinoma (11%) (cBioPortal, 2021)¹⁵¹⁻¹⁵². In HCC, FGF19 is an important driver gene^{145,153-154}, and FGF19 protein expression correlates with tumor progression and poorer prognosis¹⁵⁵. Exogenous FGF19 has been shown to promote prostate cancer tumorigenesis in a preclinical study¹⁵⁶, and the presence of FGF19-positive tissues is an independent factor for worse prognosis following radical prostatectomy¹⁵⁷.

FINDING SUMMARY

FGF19 encodes fibroblast growth factor 19, an FGFR4 ligand involved with bile acid synthesis and hepatocyte proliferation in the liver^{145,158}. FGF19 lies in a region of chromosome 11q13 frequently amplified in a diverse range of malignancies that also contains FGF3, FGF4, and CCND1¹⁵⁹. Correlation between FGF19 amplification and protein expression has been demonstrated in hepatocellular carcinoma (HCC)¹⁶⁰ but was not observed in several other tumor types¹⁵⁴.

GENOMIC FINDINGS

GENE

FGF3

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies that directly address genomic alterations in FGF₃. Inhibitors of FGF receptors, however, are undergoing clinical

trials in a number of different cancers. Limited data suggest that pan-FGFR inhibitors show activity in FGF amplified cancers; following treatment with a selective pan-FGFR inhibitor, a patient with head and neck squamous cell carcinoma (HNSCC) and amplification of 11q13 (FGF3, FGF4, FGF19) and 12p13 (FGF6 and FGF23) experienced a radiologic CR¹⁶¹.

FREQUENCY & PROGNOSIS

FGF3 lies in a region of chromosome 11q13 that also contains FGF19, FGF4, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell

cycle progression. This chromosomal region is frequently amplified in a diverse range of malignancies¹²⁴.

FINDING SUMMARY

FGF3 encodes fibroblast growth factor 3, a growth factor that plays a central role in development of the inner ear. Germline mutations in FGF3 give rise to an autosomal recessive syndrome characterized by microdontia, deafness, and complete lack of inner ear structures¹⁶².

GENE

FGF4

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

FGF4 amplification and overexpression was associated with cell sensitivity to the multikinase inhibitor sorafenib in preclinical studies¹⁶³⁻¹⁶⁴ and amplification of FGF4/FGF3 in HCC significantly correlated with patient response to sorafenib (p=0.006)¹⁶³. Limited data suggest that pan-FGFR inhibitors show activity in FGF amplified cancers; following treatment with a selective pan-FGFR

inhibitor, a patient with head and neck squamous cell carcinoma (HNSCC) and amplification of 11q13 (FGF3, FGF4, FGF19) and 12p13 (FGF6 and FGF23) experienced a radiologic CR¹⁶¹.

FREQUENCY & PROGNOSIS

FGF4 lies in a region of chromosome 11q13 that also contains FGF19, FGF3, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression. This chromosomal region is frequently amplified in a diverse range of malignancies¹²⁴ including esophageal carcinoma (35%), head and neck squamous cell carcinoma (HNSCC; 24%), breast invasive carcinoma (14%), lung squamous cell carcinoma (13%), cholangiocarcinoma (11%), bladder urothelial carcinoma (10%), stomach adenocarcinoma (7%), skin melanoma (5%), and hepatocellular carcinoma

(HCC; 5%), however FGF4 amplification is rare in hematopoietic and lymphoid malignancies, reported in less than 1% of samples analyzed (cBioPortal, 2021)¹⁵¹⁻¹⁵².

FINDING SUMMARY

FGF4 encodes fibroblast growth factor 4, which plays a central role in development of the teeth¹⁶⁵ and acts synergistically with other FGFs and SHH (sonic hedgehog) to regulate limb outgrowth in vertebrate development¹⁶⁶. FGF4 lies in a region of chromosome 11q13 that also contains FGF19, FGF3, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression. Amplification of FGF4, along with that of FGF3, FGF19, and CCND1, has been reported in a variety of cancers^{124,163,167-170} and may confer sensitivity to the multi-kinase inhibitor sorafenib¹⁶³.

GENE

KDM6A

ALTERATION

rearrangement intron 10, rearrangement exon 23

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies available to address KDM6A alterations in cancer.

FREQUENCY & PROGNOSIS

KDM6A mutations have been reported in 3.9% of samples analyzed, with the highest incidence in tumors of the urinary tract (31%), endometrium (7%), and salivary gland (6%) (COSMIC, 2021)¹⁷¹. KDM6A mutations or copy number alterations have also been identified in medulloblastoma (8.9%)¹⁷², adenoid cystic carcinoma (6.7%)¹⁷³, and metastatic prostate cancer (10%)¹⁷⁴. KDM6A inactivation has been found as a recurrent tumorigenic event in male T-cell acute lymphoblastic leukemia (T-ALL), and loss of KDM6A increased the sensitivity of T-ALL cells to therapies targeting histone H₃ lysine 27

methylation in preclinical assays¹⁷⁵. However, KDM6A overexpression has been noted in breast cancer and renal cell carcinoma, and correlated with inferior prognosis in patients with breast cancer¹⁷⁶⁻¹⁷⁸.

FINDING SUMMARY

KDM6A encodes a histone H₃ lysine 27 demethylase UTX, which functions as a transcriptional regulator¹⁷⁹. A significant number of inactivating KDM6A mutations have been found across multiple tumor types, suggesting a role as a tumor suppressor¹⁷⁹.

GENOMIC FINDINGS

GENE

NFKBIA

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies that directly target NFKBIA amplification or expression.

FREQUENCY & PROGNOSIS

In the TCGA datasets, amplification of NFKBIA has been reported with the highest incidence in lung adenocarcinoma (11.7%)⁸⁵, esophageal carcinoma (3.8%), prostate adenocarcinoma (3.4%)¹⁸⁰, lung squamous cell carcinoma (2.8%)⁸⁶, and ovarian serous cystadenocarcinoma (2.6%) (cBioPortal, 2021)¹⁵¹⁻¹⁵². Amplification or increased expression of NFKBIA in EGFR-mutant lung cancer has been reported to predict improved response to EGFR tyrosine kinase inhibitors¹⁸¹⁻¹⁸². Certain NFKBIA polymorphisms, which may affect IkBa expression levels, have been studied as risk factors for some cancer types, although the

data are mixed and conflicting183-185.

FINDING SUMMARY

NFKBIA encodes IkBa, an inhibitor of the NF-kappaB (NFkB)/REL complex. It has been reported to act as a tumor suppressor in Hodgkin's lymphoma¹⁸⁶⁻¹⁹⁰ and in glioblastoma^{183,191-192}. NFKBIA has been reported to be amplified in cancer¹⁵² and may be biologically relevant in this context¹⁹³⁻¹⁹⁴. In contrast, truncating mutations that result in loss of the majority of the IkBa protein are predicted to be inactivating.

GENE

NKX2-1

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved therapies or trials that target tumors with TTF-1 amplification or overexpression. Lung cancer cell lines that express both TTF-1 and NKX2-8, which is located in the same amplicon as NKX2-1, have demonstrated resistance to cisplatin therapy¹⁹⁵, although

conflicting data has also been reported196.

FREQUENCY & PROGNOSIS

Putative amplification of NKX2-1 has been reported with the highest incidence in lung cancer, and has been observed in 14% of adenocarcinomas⁸⁵ and 5% of squamous cell carcinomas (SCC)⁸⁶ as well as other tumor types including prostate adenocarcinomas (6%)¹⁸⁰, and poorly differentiated and anaplastic thyroid cancers (4%)¹⁹⁷. NKX2-1 mutation has been observed in 9% of acinar cell carcinomas of the pancreas¹⁹⁸, 5% of uterine carcinosarcomas¹⁹⁹, and is infrequent in other tumor types (cBioPortal, COSMIC, 2021)^{151-152,171}. TTF-1 is expressed in a majority of lung adenocarcinomas and small cell

carcinomas, as well as in a subset of thyroid and CNS tumors²⁰⁰⁻²⁰². Cytoplasmic TTF-1 expression has been reported as an adverse prognostic factor in breast carcinoma²⁰³⁻²⁰⁴. However, whether amplification and/or expression status of NKX2-1 have prognostic implications for patients with lung cancer is controversial^{195-196,205-208}. TTF-1 has been observed to have tumor-promoting as well as anti-oncogenic roles²⁰⁹⁻²¹⁰.

FINDING SUMMARY

NKX2-1 (NK2 homeobox 1) encodes the thyroid transcription factor TTF-1²¹¹. Amplification of NKX2-1 results in overexpression of TTF-1 and upregulated transcription of downstream target genes²¹².



GENOMIC FINDINGS

GENE

TP53

ALTERATION C176Y

TRANSCRIPT ID NM_000546

CODING SEQUENCE EFFECT 527G>A

VARIANT ALLELE FREQUENCY (% VAF) 39.5%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib213-216, or p53 gene therapy and immunotherapeutics such as SGT-53²¹⁷⁻²²¹ and ALT-801²²². In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% (17/ 176) and SDs in 53.4% (94/176) of patients with solid tumors; the response rate was 21.1% (4/19) for patients with TP53 mutations versus 12.1% (4/ 33) for patients who were TP53 wild-type223. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 31.9% (30/ 94, 3 CR) ORR and a 73.4% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer²²⁴. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 42.9% (9/21, 1 CR) ORR and a 76.2% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer²²⁵. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone²²⁶. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/ or recurrent gastric cancer experienced a 24.0% (6/25) ORR with adavosertib combined with paclitaxel²²⁷. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71.4% (5/7)

response rate for patients with TP53 alterations²²⁸. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75.0% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage²²¹. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53-mutated, but not TP53-wildtype, breast cancer xenotransplant mouse model²²⁹. Missense mutations leading to TP₅₃ inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246²³⁰⁻²³². In a Phase 1b trial for patients with p53-positive highgrade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR²³³. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies²³⁴⁻²³⁵; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies²³⁶⁻²³⁷. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer; mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)85-86,238-243, including 42-52% of lung adenocarcinomas and 58-83% of lung squamous cell carcinomas (cBioPortal, COSMIC, Feb 2021)48-49,85-86. TP53 homozygous deletion has been observed in 1.4% of lung adenocarcinoma and <1% of lung squamous cell carcinoma cases (cBioPortal, Feb 2021)151-152. In one study of 55 patients with lung adenocarcinoma, TP53 alterations correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors pembrolizumab and nivolumab in this study²⁴⁴. Mutations in TP₅₃ have been associated with lymph node metastasis in patients with lung adenocarcinoma245.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers²⁴⁶. Alterations such as seen here may disrupt TP53 function or expression²⁴⁷⁻²⁵¹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Sep 2021) 252 . Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers²⁵³⁻²⁵⁵, including sarcomas²⁵⁶⁻²⁵⁷. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000²⁵⁸ to 1:20,000²⁵⁷. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30²⁵⁹. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion260-265. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy²⁶⁰⁻²⁶¹. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease²⁶⁶. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to $\mathrm{CH}^{264,267\text{-}268}.$ Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Afatinib

Assay findings association

EGFR

exon 19 deletion (T751_I759>N)

AREAS OF THERAPEUTIC USE

Afatinib is an irreversible kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4. It is available in the EU to treat patients with advanced nonsmall cell lung cancer (NSCLC) and activating EGFR mutations and for the treatment of patients with advanced squamous NSCLC after progression on platinum-based chemotherapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to a fatinib or dacomitinib for patients with non-small cell lung cancer $^{66\text{-}67,269\text{-}270}$, whereas data for patients with other tumor types are limited $^{69\text{-}74,271}$.

SUPPORTING DATA

Afatinib has shown significant clinical activity for patients with NSCLC and the EGFR common sensitizing mutations L858R or exon 19 deletions, based on extensive clinical evidence^{66,269,272-275}. Two randomized Phase 3 trials reported significantly improved median PFS from afatinib compared with chemotherapy for patients with EGFR common sensitizing mutations (LUX-Lung 3, 13.6 vs. 6.9 months, HR 0.47, p<0.001; LUX-Lung 6, 11.0 vs. 5.6 months, HR 0.28, p<0.0001)66,269. However, while afatinib significantly increased OS relative to chemotherapy for patients with EGFR exon 19 alterations in these two trials (LUX-Lung 3, 33.3 vs. 21.1 months, HR=0.54; LUX-Lung 6, 31.4 vs. 18.4 months, HR=0.64), no significant OS differences were observed in treatment for patients with L858R mutation¹⁰³. A similar alterationspecific difference was observed for EGFR-mutated treatment-naive NSCLC in a retrospective analysis, which reported numerically longer median OS from secondversus first-generation EGFR TKIs (48.8 vs. 26.4 months, HR=0.59) for patients with exon 19 deletions, but no substantial difference for patients with L858R (25.4 vs. 20.6 months, HR=0.90)²⁷². A Phase 2b study of first-line afatinib compared with gefitinib, also for NSCLC with exon 19 deletions or L858R, reported similar median OS for the two therapies (27.9 vs. 24.5 months, HR=0.86) but significantly longer time-to-treatment-failure (13.7 vs. 11.5 months, HR=0.75) and higher ORR (73% vs. 56%, p=0.0018) with afatinib²⁷³. Patients with metastatic

NSCLC and common EGFR mutations who progressed on prior chemotherapy experienced an ORR of 50.0% (30/ 60) from afatinib in a Phase 4 trial²⁷⁴. As first-line therapy for NSCLC with EGFR exon 19 deletions or L858R, prospective or randomized Phase 2 trials have reported a median PFS of 10.2 months and OS of 24.8 months for patients unfit for chemotherapy²⁷⁵ and an ORR of 72.5% (n=40, 1 CR), DCR of 100% (40/40), and median PFS and OS of 15.2 and 30.0 months, respectively, for elderly patients ≥70 years old²⁷⁶. A retrospective study of afatinib administered to Asian patients with NSCLC, 99% of whom were previously treated with erlotinib and/or gefitinib, reported an ORR of 27.4% (63/230) for patients with common sensitizing EGFR mutations and an ORR of 24.4% (105/431) for the entire cohort²⁷⁷. In a case report, a patient with NSCLC with exon 19 deletion and leptomeningeal metastases experienced an ongoing 16-month PR from afatinib in extracranial, brain, and leptomeningeal lesions²⁷⁸. For patients with erlotinib- or gefitinib-resistant NSCLC and EGFR mutations, Phase 2/ 3 studies of afatinib treatment have generally reported ORRs of only 7 to 9%²⁷⁹⁻²⁸⁴; however, DCRs of more than 50% have been observed²⁸³. In a Phase 1b or observational study, patients with EGFR-mutated NSCLC who progressed on afatinib experienced further clinical benefit from subsequent treatment with afatinib and cetuximab²⁸⁵ or osimertinib²⁸⁶, respectively. Extensive clinical data have demonstrated that afatinib is effective for patients with EGFR-mutated advanced NSCLC, including exon 19 deletions and L858 mutations, as well as uncommon sensitizing mutations in exons 18 or $20^{66,103,269,273,275,277,287}$. A fatinib has also shown activity for patients with advanced NSCLC and ERBB2 mutations, most of which were exon 20 insertions $^{283,288\text{-}298}$. The randomized Phase 3 LUX-Lung 8 trial comparing afatinib with erlotinib as second-line therapy for advanced lung squamous cell carcinoma (SCC) reported significantly longer median OS (7.9 vs. 6.8 months, HR=0.81), significantly longer median PFS (2.6 vs. 1.9 months, HR=0.81), and higher DCR (51% vs. 40%, p=0.002) for patients treated with afatinib²⁸⁷. For patients who progressed on afatinib monotherapy, additional clinical benefit has been reported from afatinib combined with paclitaxel²⁹⁹.

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Dacomitinib

Assay findings association

EGFR

exon 19 deletion (T751_I759>N)

AREAS OF THERAPEUTIC USE

Dacomitinib is a second-generation irreversible tyrosine kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4/HER4. It is available in the EU for first-line treatment of patients with advanced nonsmall cell lung cancer (NSCLC) with EGFR activating mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer^{66-67,269-270}, whereas data for patients with other tumor types are limited^{69-74,271}. Patients with untreated advanced NSCLC and EGFR exon 19 deletions achieved an ORR of $76\%^{105}$ and a median OS of 34.1 months with dacomitinib⁶⁷.

SUPPORTING DATA

A randomized Phase 3 trial in patients with NSCLC with activating EGFR mutations (primarily L858R or exon 19 deletions) reported improved clinical benefit with first-line dacomitinib compared with gefitinib (median OS,

34.1 vs. 26.8 months, HR=0.760; median PFS, 14.7 vs. 9.2 months, HR=0.59)105,300; median OS was 34.1 to 36.7 months and ORR was 74.9% to 79.3%, depending on the dosing regimen³⁰¹. A pooled subgroup analysis of patients with NSCLC with activating EGFR mutations reported improved clinical efficacy with dacomitinib treatment compared with erlotinib (median PFS, 14.6 vs, 9.6 months, HR=0.717; median OS, 26.6 vs, 23.2 months, HR=0.737)³⁰². Reduced efficacy of dacomitinib treatment in patients with NSCLC harboring the EGFR T790M mutation has been reported in multiple studies³⁰³⁻³⁰⁵. A Phase 1 trial of combination dacomitinib and a MEK1/2 inhibitor for patients with KRAS-mutated CRC, NSCLC, or pancreatic cancer reported 20/36 SDs and 16 PDs, however toxicity from this combination prevented longterm treatment in this patient population³⁰⁶. A Phase 2 study of dacomitinib in patients with NSCLC who had been previously treated with chemotherapy or erlotinib and were not selected for EGFR mutations reported an ORR of 4.5% (3/66)304. In one study, the combination of dacomitinib and crizotinib was ineffective and associated with high toxicity in patients with NSCLC³⁰⁷.

Erlotinib

Assay findings association

ECED

exon 19 deletion (T751_I759>N)

AREAS OF THERAPEUTIC USE

Erlotinib is an EGFR tyrosine kinase inhibitor. It is available in the EU to treat advanced non-small cell lung cancer (NSCLC) as first-line therapy or switch maintenance therapy for patients with EGFR-activating mutations and as second-line therapy for patients who have progressed on prior chemotherapy. Erlotinib is also available in combination with gemcitabine to treat metastatic pancreatic cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression^{64,308-310}.

SUPPORTING DATA

For patients with EGFR-mutated NSCLC, the Phase 3 EURTAC trial reported improved PFS with first-line erlotinib relative to platinum-based chemotherapy (9.7 vs. 5.2 months, HR=0.37)⁶⁴. A Phase 3 study reported similar efficacy of erlotinib and gefitinib for patients with EGFR-mutated NSCLC³¹¹. Meta-analysis of studies comparing erlotinib or gefitinib versus chemotherapy in the first-line setting reported no significant improvement in OS for patients with EGFR-mutated NSCLC; however, the lack of

improved OS was attributed to the effectiveness of postprogression salvage therapy³¹². In the maintenance setting, the placebo-controlled Phase 3 SATURN trial reported significantly improved PFS with maintenance erlotinib following first-line platinum-based chemotherapy irrespective of EGFR status; however, the largest effect was seen for patients with EGFR mutations $(HR=0.10)^{308}$. In the neoadjuvant setting, a Phase 2 trial reported a numerically improved ORR and significantly longer PFS with erlotinib compared with chemotherapy for patients with advanced EGFR-mutated NSCLC 309 . In the placebo-controlled Phase 3 RELAY trial, the addition of ramucirumab to erlotinib improved PFS for previously untreated patients with NSCLC harboring EGFR L858R or exon 19 deletion (19.4 vs. 12.4 months, HR=0.59)313. In a Phase 2 trial, no clinical benefit was observed from the addition of bevacizumab to erlotinib for patients with NSCLC harboring EGFR exon 19 deletion or L858R mutation³¹⁴. In one study, median PFS (4.1 vs. 11.7 months, HR=9.7) and median OS (14.1 vs. 47.0 months, HR=10.2) were significantly shorter for patients with NSCLC harboring EGFR L747_A750>P (n=6) relative to those with deletions affecting EGFR E746_A750 (n=24) treated with first-line erlotinib315. The Phase 3 BR.21 trial demonstrated prolonged OS for genomically unselected patients with NSCLC treated with erlotinib compared with those treated with standard chemotherapy316.





THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Gefitinib

Assay findings association

EGFR

exon 19 deletion (T751_I759>N)

AREAS OF THERAPEUTIC USE

Gefitinib is an EGFR tyrosine kinase inhibitor available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) with activating EGFR mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy^{310,317-322}, and responses have been reported for patients with EGFR-rearranged NSCLC³²³⁻³²⁴.

SUPPORTING DATA

A Phase 3 trial of first-line gefitinib therapy for patients with NSCLC and EGFR exon 19 deletions or L858R mutations reported a longer PFS (9.2 months vs. 6.3 months)³¹⁹ but no change in median OS (34.9 months vs. 37.2 months) compared with patients treated with cisplatin plus docetaxel (median OS of 37.2 months)³²⁵. Gefitinib achieved an ORR of 69.8% and an OS of 19.2 months as first-line treatment for Caucasian patients with non-small cell lung carcinoma (NSCLC) and EGFR sensitizing mutations⁶⁵. In the retrospective analysis of a Phase 3 study for East Asian patients, gefitinib was

reported to have a longer PFS for patients with EGFR mutation-positive NSCLC compared with carboplatin/ paclitaxel doublet chemotherapy 320,326 . Two Phase 3 trials of gefitinib plus pemetrexed and carboplatin compared with gefitinib alone for patients with advanced NSCLC harboring EGFR activating mutations reported significantly higher ORRs (75.3% and 84% vs. 62.5% and 67%), longer median PFSs (16 and 20.9 months vs. 8 and 11.9 months), and longer median OSs (50.9 months and not reached vs. 17 and 38.8 months) with combination treatment; however, combination treatment was associated with increased Grade 3 or higher adverse events³²⁷⁻³²⁸. Retrospective analysis of East Asian patients with advanced NSCLC receiving first-line gefitinib therapy reported that patients with EGFR exon 19 mutations experienced a longer median PFS (10.9 months) compared with patients with EGFR mutations in exon 18 (7.9 months), exon 20 (1.2 months), exon 21 (7.7 months), or double mutations (5.7 months); however, no differences in OS were seen between EGFR mutations³²⁹. In a Phase 1 study for treatment-naive patients with NSCLC, best ORRs of 78% (7/9) were observed in patients treated with combination gefitinib and the PD-L1 inhibitor durvalumab as first-line treatment and of 80% (8/10) in those treated with the combination after gefitinib monotherapy³³⁰.



THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Osimertinib

Assay findings association

EGFR

exon 19 deletion (T751_I759>N)

AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR tyrosine kinase inhibitor (TKI) that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is available in the EU in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors harbor EGFR T790M mutations or activating mutations, including EGFR exon 19 deletions and exon 21 L858R mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/ or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer^{68,104,323,331-332}. Patients with untreated advanced NSCLC and EGFR exon 19 deletions or L858R mutations achieved an ORR of 80% and a median PFS of 21.4 and 14.4 months, respectively¹⁰⁴.

SUPPORTING DATA

The Phase 3 FLAURA study reported that, relative to erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (18.9 vs. 10.2 months, HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80)

for patients with advanced NSCLC and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858)104,333. In the Phase 3 ADAURA study, patients with early Stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer PFSs on osimertinib compared to placebo in the adjuvant setting (not reached vs. 28.1 months; HR=0.21)334. A Phase 1 study reported that T790M-negative patients with acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months⁶⁸. A Phase 1b/2 study evaluating osimertinib in combination with the CD₇₃ inhibitor oleclumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/19), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 1/2 trial of osimertinib in combination with bevacizumab for patients with untreated metastatic EGFR-mutated non-small cell lung cancer (NSCLC) reported an 80% (39/49) ORR, a 100% (6/6, 2 CRs) central nervous system response rate, median PFS of 19 months, and a 1-year PFS rate of 72%335. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively³³⁶.

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies listed in this report may not be complete and exhaustive and the therapeutic agents are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type.





CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized in the following descending order: Pediatric trial qualification → Geographical proximity → Trial phase → Trial verification within last 2 months. While every effort is made to ensure the accuracy of the information contained below, the

information available in the public domain is continually updated and should be investigated by the physician or research staff. The clinical trials listed in this report may not be complete and exhaustive or may include trials for which the patient does not meet the clinical trial

enrollment criteria. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov or local registries in your region.

CCND1

RATIONALE

CCND1 amplification or overexpression may activate CDK4/6 and may predict sensitivity to

single-agent CDK4/6 inhibitors.

auteration amplification

NCT04000529 PHASE 1

Phase Ib Study of TNO155 in Combination With Spartalizumab or Ribociclib in Selected Malignancies

TARGETS
PD-1, SHP2, CDK6, CDK4

LOCATIONS: Barcelona (Spain), Koeln (Germany), Bruxelles (Belgium), Massachusetts, Chengdu (China), Hong Kong (Hong Kong), Singapore (Singapore), Chuo ku (Japan), Westmead (Australia)

NCT02664935 PHASE 2

National Lung Matrix Trial: Multi-drug Phase II Trial in Non-Small Cell Lung Cancer TARGETS

FGFRs, mTORC1, mTORC2, CDK4, CDK6, ALK, AXL, MET, ROS1, TRKA, TRKC, MEK, AKTs, EGFR, PD-L1, DDR2, FLT3, KIT, PDGFRA, RET, TRKB, VEGFRs

LOCATIONS: Maidstone (United Kingdom), Colchester (United Kingdom), London (United Kingdom), Southampton (United Kingdom), Cambridge (United Kingdom), Oxford (United Kingdom), Exeter (United Kingdom), Bristol (United Kingdom), Leicester (United Kingdom), Cardiff (United Kingdom)

NCTO4801966

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS

CDK4, CDK6, PI3K-alpha, PD-L1, MEK,

LOCATIONS: Melbourne (Australia)

NCT04553133 PHASE 2

PF-07104091 as a Single Agent and in Combination Therapy TARGETS

Aromatase, CDK4, CDK6, CDK2

LOCATIONS: Massachusetts, Michigan, Texas

NCT02896335 PHASE 2

Palbociclib In Progressive Brain Metastases TARGETS

CDK4, CDK6

PARP, PD-1, BRAF

LOCATIONS: Massachusetts



CLINICAL TRIALS

NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO

LOCATIONS: Montreal (Canada), Ottawa (Canada), Kingston (Canada), Toronto (Canada), London (Canada), Saskatoon (Canada), Regina (Canada), Edmonton (Canada), Vancouver (Canada)

NCT03065062	PHASE 1
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors	TARGETS PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6
LOCATIONS: Massachusetts	
NCT03454035	PHASE 1
Ulixertinib/Palbociclib in Patients With Advanced Pancreatic and Other Solid Tumors	TARGETS MAPK3, MAPK1, CDK4, CDK6

LOCATIONS: North Carolina

NCT02897375		PHASE 1	
Palbociclib With Cisplatin or Carboplatin in Advanced Solid Tumors		TARGETS CDK4, CDK6	
LOCATIONS: Georgia			





CLINICAL TRIALS

CDK4

RATIONALE

CDK4 amplification may predict sensitivity to

CDK₄/6 inhibitors.

ALTERATION amplification - equivocal

NCT03099174

This Study in Patients With Different Types of Cancer (Solid Tumours) Aims to Find a Safe Dose of Xentuzumab in Combination With Abemaciclib With or Without Hormonal Therapies. The Study Also Tests How Effective These Medicines Are in Patients With Lung and Breast Cancer.

PHASE 1

TARGETS

CDK4, CDK6, IGF-1, IGF-2, Aromatase, ER

LOCATIONS: Paris (France), Marseille (France), Barcelona (Spain), L'Hospitalet de Llobregat (Spain), Besançon (France), Madrid (Spain), Pozuelo de Alarcón (Spain), Malaga (Spain), Plerin Sur Mer (France), København Ø (Denmark)

NCT04000529

Phase Ib Study of TNO155 in Combination With Spartalizumab or Ribociclib in Selected Malignancies
TARGETS
PD-1, SHP2, CDK6, CDK4

LOCATIONS: Barcelona (Spain), Koeln (Germany), Bruxelles (Belgium), Massachusetts, Chengdu (China), Hong Kong (Hong Kong), Singapore (Singapore), Chuo ku (Japan), Westmead (Australia)

National Lung Matrix Trial: Multi-drug Phase II Trial in Non-Small Cell Lung Cancer

TARGETS
FGFRs, mTORC1, mTORC2, CDK4,
CDK6, ALK, AXL, MET, ROS1, TRKA,
TRKC, MEK, AKTs, EGFR, PD-L1, DDR2,
FLT3, KIT, PDGFRA, RET, TRKB, VEGFRS

LOCATIONS: Maidstone (United Kingdom), Colchester (United Kingdom), London (United Kingdom), Southampton (United Kingdom), Cambridge (United Kingdom), Oxford (United Kingdom), Exeter (United Kingdom), Bristol (United Kingdom), Leicester (United Kingdom), Cardiff (United Kingdom)

NCTO4801966

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS

CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

TARGETS

VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, CDK4, CDK6, CSF1R, FLT3, KIT, RET, mTOR, EGFR, ERBB2, ERBB3, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4



CLINICAL TRIALS

NCT03994796	PHASE 2
Genetic Testing in Guiding Treatment for Patients With Brain Metastases	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR
LOCATIONS: Massachusetts, Vermont, New Hampshire, New York, Connecticut, New Jersey	
NCTO4553133	PHASE 2
PF-07104091 as a Single Agent and in Combination Therapy	TARGETS Aromatase, CDK4, CDK6, CDK2
LOCATIONS: Massachusetts, Michigan, Texas	
NCT02896335	PHASE 2
Palbociclib In Progressive Brain Metastases	TARGETS CDK4, CDK6
LOCATIONS: Massachusetts	
NCT03310879	PHASE 2
Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6	TARGETS CDK4, CDK6
LOCATIONS: Massachusetts	
NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO
LOCATIONS: Montreal (Canada), Ottawa (Canada), Kingston (Canada), Toronto (Canada), London (Canada), Vancouver (Canada)	anada), Saskatoon (Canada), Regina (Canada),



CLINICAL TRIALS

GENE EGFR

EGFR ALTERATION

exon 19 deletion (T751 1759>N)

RATIONALE

EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome

resistance to current agents include nextgeneration EGFR inhibitors and combination therapies.

NCT04487080 PHASE 3

A Study of Amivantamab and Lazertinib Combination Therapy Versus Osimertinib in Locally
Advanced or Metastatic Non-Small Cell Lung Cancer

TARGETS
MET, EGFR

LOCATIONS: Catania (Italy), Napoli (Italy), Avellino (Italy), Meldola (Italy), Ravenna (Italy), Rozzano (Italy), Milano (Italy), Marseille Cedex 20 (France), Monza (Italy), Orbassano (Italy)

NCT04811001 PHASE 2

Best EGFR-TKI Sequence in NSCLC Harboring EGFR Mutations

TARGETS
EGFR, ERBB2, ERBB4

LOCATIONS: Napoli (Italy), Roma (Italy), Padova (Italy), Novara (Italy)

NCT04077463 PHASE 1

A Study of Lazertinib as Monotherapy or in Combination With JNJ-61186372 in Japanese Participants
With Advanced Non-small Cell Lung Cancer

TARGETS
EGFR, MET

LOCATIONS: Napoli (Italy), Ravenna (Italy), Marseille (France), Milano (Italy), Barcelona (Spain), Lyon Cedex 8 (France), Gauting (Germany), Stuttgart (Germany), Bordeaux (France), Poitiers (France)

NCT04248829 PHASE 3

Clinical Trial of YH25448(Lazertinib) as the First-line Treatment in Patients With EGFR Mutation
Positive Locally Advanced or Metastatic NSCLC (LASER301)

TARGETS
EGFR

LOCATIONS: Patras (Greece), Larissa (Greece), Thessaloníki (Greece), Athens (Greece), Kragujevac (Serbia), Niš (Serbia), Belgrade (Serbia), Sremska Kamenica (Serbia), Székesfehérvár (Hungary), Tatabánya (Hungary)

NCT03521154 PHASE 3

A Global Study to Assess the Effects of Osimertinib Following Chemoradiation in Patients With Stage
III Unresectable Non-small Cell Lung Cancer (LAURA)

TARGETS
EGFR

LOCATIONS: Pécs (Hungary), Barcelona (Spain), Székesfehérvár (Hungary), Törökbálint (Hungary), Budapest (Hungary), Valencia (Spain), Izmir (Turkey), Gyöngyös - Mátraháza (Hungary), Istanbul (Turkey), San Sebastián (Spain)

NCTO4035486 PHASE 3

A Study of Osimertinib With or Without Chemotherapy as 1st Line Treatment in Patients With

Mutated Epidermal Growth Factor Receptor Non-Small Cell Lung Cancer (FLAURA2)

TARGETS

EGFR

LOCATIONS: Montpellier (France), Lyon (France), Banska Bystrica (Slovakia), Olomouc (Czechia), Praha (Czechia), Praha 5 (Czechia), Poprad (Slovakia), Kosice (Slovakia), Ostrava - Vitkovice (Czechia), Bordeaux Cedex (France)



CLINICAL TRIALS

NCT04413201	PHASE 4
AFAMOSI: Efficacy and Safety of Afatinib Followed by Osimertinib Compared to Osimertinib in Patients With EGFRmutated/T790M Mutation Negative Nonsquamous NSCLC	TARGETS EGFR, ERBB2, ERBB4

LOCATIONS: Immenstadt (Germany), Konstanz (Germany), Regensburg (Germany), Löwenstein (Germany), Hamm (Germany), Berlin (Germany), Bremen (Germany), Hamburg (Germany)

NCT03865511	PHASE 2	
MEchanisms of Resistance in EGFR Mutated Nonpretreated Advanced Lung Cancer Receiving OSimErtib	TARGETS EGFR	

LOCATIONS: Toulon (France), Cholet (France), Le Mans (France), Nantes (France)

NCT04233021	PHASE 2	
Study of Osimertinib in Patients With a Lung Cancer With Brain or Leptomeningeal Metastases With EGFR Mutation	TARGETS EGFR	

LOCATIONS: Toulon (France), Marseille (France), Aix-en-Provence (France), Avignon (France), Montpellier (France), Grenoble (France), Pierre-Bénite (France), Lyon (France), Villefranche-sur-Saône (France), Toulouse (France)

NCT04619004		PHASE 2
HERTHENA-Lung01: Patritumab Deruxtecan in Subjects With I mutated Non-Small Cell Lung Cancer	Metastatic or Locally Advanced EGFR-	TARGETS ERBB3

LOCATIONS: Rozzano (Italy), Milano (Italy), Orbassano (Italy), Grenoble (France), Barcelona (Spain), Lyon (France), Toulouse (France), Zaragoza (Spain), Dresden (Germany), Villejuif (France)





APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ATR

E471Q

IKZF1 amplification

CREBBP P323R

IRS2 A701_V702insAA **EGFR** amplification

MED12 Q2119_Q2120insHQQQ **FAM123B** F127L

RAC1 amplification



FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	ЕРНА3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF11	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

^{*}TERC is an NCRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER GENOMIC SIGNATURES

Loss of Heterozygosity (LOH) score Microsatellite (MS) status

Tumor Mutational Burden (TMB)

^{**}Promoter region of TERT is interrogated

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

ABOUT FOUNDATIONONE CDX

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffinembedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes. Using an Illumina® HiSeq platform, hybrid

capture–selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g.,gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: The association of a therapy with a genomic alteration or signature does not necessarily indicate pharmacologic effectiveness (or lack thereof); no association of a therapy with a genomic alteration or signature does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness).

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials Ranking of Therapies in Summary Table
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK* (NCCN*) CATEGORIZATION

Genomic signatures and gene alterations detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each genomic signature or gene alteration. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X,

APPENDIX

About FoundationOne®CDx

- "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- 2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/ pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
- **3.** The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding armand chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- 4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- 5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- **6.** Reflex testing to an alternative FDA approved

companion diagnostic should be performed for patients who have an ERBB2 amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of ERBB2 copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in https://www.mycancergenome.org/content/ disease/breast-cancer/ERBB2/238/ report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
INDELS Repeatability	%CV*

^{*}Interquartile Range = 1^{st} Quartile to 3^{rd} Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of followup germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MLH1, MSH2, MSH6, MUTYH, PALB2, PMS2, POLE, RAD51C, RAD51D, RET, SDHA, SDHB, SDHC, SDHD, TSC2, and VHL, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are ASXL1, CBL, DNMT3A, IDH2, JAK2, KMT2D (MLL2), MPL, MYD88, SF3B1, TET2, and U2AF1 and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively

APPENDIX

About FoundationOne®CDx

determine if this alteration is present in tumor or is **SELECT ABBREVIATIONS** secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE **RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS					
ABBREVIATION	DEFINITION				
CR	Complete response				
DCR	Disease control rate				
DNMT	DNA methyltransferase				
HR	Hazard ratio				
ITD	Internal tandem duplication				
MMR	Mismatch repair				
muts/Mb	Mutations per megabase				
NOS	Not otherwise specified				
ORR	Objective response rate				
os	Overall survival				
PD	Progressive disease				
PFS	Progression-free survival				
PR	Partial response				
SD	Stable disease				
ткі	Tyrosine kinase inhibitor				

MR Suite Version 5.2.0

The median exon coverage for this sample is 943x

29436178

9329646

APPENDIX

References

- 1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
- 2. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
- 3. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
- 4. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
- 5. Ayers et al., 2016; ASCO-SITC Abstract P60
- 6. Warth A, et al. Virchows Arch. (2016) pmid: 26637197
- 7. Ninomiya H, et al. Br. J. Cancer (2006) pmid: 16641899 8. Vanderwalde A, et al. Cancer Med (2018) pmid:
- 9. Zang YS, et al. Cancer Med (2019) pmid: 31270941
- 10. Dudley JC, et al. Clin. Cancer Res. (2016) pmid: 26880610
- 11. Takamochi K, et al. Lung Cancer (2017) pmid: 28676214 Pylkkänen L, et al. Environ. Mol. Mutagen. (1997) pmid:
- 13. Gonzalez R, et al. Ann. Oncol. (2000) pmid: 11061602
- 14. Chen XQ, et al. Nat. Med. (1996) pmid: 8782463
- 15. Merlo A, et al. Cancer Res. (1994) pmid: 8174113
- 16. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
- 17. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
- 18. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
- 19. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
- 20. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
- 21. Boland CR, et al. Gastroenterology (2010) pmid: 20420947
- 22. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
- 23. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
- 24. Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
- 25. Cristescu R. et al. Science (2018) pmid: 30309915
- 26. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
- 27. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid:
- 28. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
- 29. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
- 30. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
- 31. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
- 32. Rizvi NA, et al. Science (2015) pmid: 25765070
- 33. Colli LM, et al. Cancer Res. (2016) pmid: 27197178
- 34. Wang VE, et al. J Immunother Cancer (2017) pmid:
- 35. Carbone DP, et al. N. Engl. J. Med. (2017) pmid: 28636851
- 36. Rizvi H, et al. J. Clin. Oncol. (2018) pmid: 29337640
- 37. Forde PM, et al. N. Engl. J. Med. (2018) pmid: 29658848
- 38. Miao D, et al. Nat. Genet. (2018) pmid: 30150660
- **39.** Chae YK, et al. Clin Lung Cancer (2019) pmid: 30425022
- 40. Paz-Ares et al., 2019; ESMO Abstract LBA80
- 41. Hellmann MD, et al. N. Engl. J. Med. (2019) pmid:
- 42. Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
- 43. Spigel et al., 2016; ASCO Abstract 9017
- 44. Xiao D, et al. Oncotarget (2016) pmid: 27009843
- 45. Shim HS, et al. J Thorac Oncol (2015) pmid: 26200269
- 46. Govindan R, et al. Cell (2012) pmid: 22980976
- **47.** Ding L, et al. Nature (2008) pmid: 18948947 48. Imielinski M, et al. Cell (2012) pmid: 22980975
- 49. Kim Y, et al. J. Clin. Oncol. (2014) pmid: 24323028
- 50. Stein et al., 2019; DOI: 10.1200/PO.18.00376
- 51. Chen Y, et al. J. Exp. Clin. Cancer Res. (2019) pmid: 31088500

- **52.** Yu H, et al. J Thorac Oncol (2019) pmid: 30253973
- 53. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
- 54. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
- 55. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
- **56.** Johnson BE, et al. Science (2014) pmid: 24336570 57. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
- 58. Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
- **59.** Briggs S, et al. J. Pathol. (2013) pmid: 23447401
- 60. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
- 61. Nature (2012) pmid: 22810696
- 62. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
- 63. Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
- 64. Rosell R, et al. Lancet Oncol. (2012) pmid: 22285168
- 65. Douillard JY, et al. Br. J. Cancer (2014) pmid: 24263064
- 66. Sequist LV, et al. J. Clin. Oncol. (2013) pmid: 23816960
- 67. Mok TS, et al. J. Clin. Oncol. (2018) pmid: 29864379 68. Jänne PA, et al. N. Engl. J. Med. (2015) pmid: 25923549
- 69. Hong MH, et al. Cancer (2020) pmid: 32749686
- 70. Kim HS, et al. Oncotarget (2015) pmid: 26462025
- 71. Kim HS, et al. Clin. Cancer Res. (2015) pmid: 25424851
- 72. Mondal G, et al. Acta Neuropathol (2020) pmid: 32303840
- 73. Cavalieri S, et al. Eur. J. Cancer (2018) pmid: 29734047
- 74. Chi AS, et al. JCO Precis Oncol (2020) pmid: 32923886
- 75. Leighl et al., 2021; ESMO Abstract 1192MO
- 76. Cho et al., 2020; ESMO Abstract 12580
- 77. Bauml et al., 2021; ASCO Abstract 9006
- 78. Shu et al., 2021; ESMO Abstract 1193MO
- 79. Janne et al., 2021; ASCO Abstract 9007
- 80. Ahn MJ, et al. Lancet Respir Med (2017) pmid: 29056570
- 81. Yang Z, et al. Sci Transl Med (2016) pmid: 27928026
- 82. Ahn MJ, et al. Lancet Oncol (2019) pmid: 31587882
- 83. Socinski MA, et al. N. Engl. J. Med. (2018) pmid: 29863955
- 84. Vallee A, et al. Int. J. Oncol. (2013) pmid: 23934203
- 85. Nature (2014) pmid: 25079552
- 86. Nature (2012) pmid: 22960745
- 87. Watzka SB, et al. Eur J Cardiothorac Surg (2010) pmid: 20353893
- 88. Liang Z, et al. BMC Cancer (2010) pmid: 20637128
- 89. Grob TJ, et al. Lung Cancer (2013) pmid: 23238037
- 90. Park S, et al. Histol. Histopathol. (2012) pmid: 22207554
- 91. Dobashi Y, et al. Hum. Pathol. (2011) pmid: 21040950
- 92. Ludovini V, et al. Cancer Chemother. Pharmacol. (2013) pmid: 23314677
- 93. Skrzypski M, et al. Clin Lung Cancer (2013) pmid: 23870818
- 94. Kim SH, et al. Histol. Histopathol. (2012) pmid: 22419022
- 95. Lee JS, et al. Ann. Surg. Oncol. (2013) pmid: 23525704 96. Oakley GJ, et al. J Thorac Oncol (2011) pmid: 21587084
- 97. Marks JL, et al. J Thorac Oncol (2008) pmid: 18303429
- 98. Izar B, et al. Ann. Thorac. Surg. (2013) pmid: 23932319 99. Ciardiello F, et al. N. Engl. J. Med. (2008) pmid:
- 100. Lynch TJ, et al. N. Engl. J. Med. (2004) pmid: 15118073
- 101. Paez JG, et al. Science (2004) pmid: 15118125

18337605

- 102. Pao W, et al. Proc. Natl. Acad. Sci. U.S.A. (2004) pmid:
- 103. Yang JC, et al. Lancet Oncol. (2015) pmid: 25589191
- 104. Soria JC, et al. N. Engl. J. Med. (2018) pmid: 29151359

- 105. Wu YL, et al. Lancet Oncol. (2017) pmid: 28958502
- 106. Gilmer TM, et al. Cancer Res. (2008) pmid: 18199554
- 107. Foster SA, et al. Cancer Cell (2016) pmid: 26996308
- 108. Morschhauser F, et al. Haematologica (2020) pmid:
- 109. Flaherty KT, et al. Clin. Cancer Res. (2012) pmid: 22090362
- 110. Finn RS, et al. Lancet Oncol. (2015) pmid: 25524798
- 111. Infante JR, et al. Clin. Cancer Res. (2016) pmid: 27542767
- 112. Patnaik A, et al. Cancer Discov (2016) pmid: 27217383
- 113. Leonard JP, et al. Blood (2012) pmid: 22383795
- 114. Dickler MN, et al. Clin. Cancer Res. (2017) pmid: 28533223
- 115. Clark et al., 2019; AACR Abstract LB-010/2
- 116. Peguero et al., 2016; ASCO Abstract 2528
- 117. Chen Y. et al. Front Immunol (2020) pmid: 32903763
- 118. Litchfield K, et al. Cell (2021) pmid: 33508232
- 119. Reissmann PT, et al. J. Cancer Res. Clin. Oncol. (1999) pmid: 10190311
- 120. Marchetti A, et al. Int. J. Cancer (1998) pmid: 9462706
- 121. Sun W, et al. J Biomed Res (2013) pmid: 23720678
- 122. Gautschi O, et al. Lung Cancer (2007) pmid: 17070615
- 123. Elsheikh S, et al. Breast Cancer Res. Treat. (2008) pmid: 17653856
- 124. Fu M, et al. Endocrinology (2004) pmid: 15331580
- 125. Takahashi-Yanaga F, et al. Cell. Signal. (2008) pmid: 18023328
- 126. Dickson MA, et al. J. Clin. Oncol. (2013) pmid: 23569312
- 127. Dickson et al., 2019: ASCO Abstract 11004
- 128. Dickson MA, et al. JAMA Oncol (2016) pmid: 27124835
- 129. Campbell JD. et al. Nat. Genet. (2016) pmid: 27158780
- 130. Wikman H, et al. Genes Chromosomes Cancer (2005) pmid: 15543620
- 131. Borczuk AC, et al. Am. J. Pathol. (2003) pmid: 14578194
- 132. Wu A, et al. J Transl Med (2011) pmid: 21477379
- 133. Puyol M, et al. Cancer Cell (2010) pmid: 20609353
- 134. Choi YJ, et al. Oncogene (2014) pmid: 23644662
- 135. Cell (1995) pmid: 7736585 136. Musgrove EA, et al. Nat. Rev. Cancer (2011) pmid:
- 21734724
- 137. Rao SK, et al. J. Neurooncol. (2010) pmid: 19609742 138. Chung L, et al. Am. J. Surg. Pathol. (2009) pmid:
- 19574885 139. Ragazzini P, et al. Histol. Histopathol. (2004) pmid:
- 140. Dujardin F, et al. Mod. Pathol. (2011) pmid: 21336260
- 141. Zhang K, et al. Cancer Res. (2013) pmid: 23393200
- 142. Horvai AE, et al. Mod. Pathol. (2009) pmid: 19734852 143. Guagnano V, et al. Cancer Discov (2012) pmid:
- 23002168 144. Hatlen MA, et al. Cancer Discov (2019) pmid: 31575540
- 145. Hagel M, et al. Cancer Discov (2015) pmid: 25776529
- 146. Kim RD, et al. Cancer Discov (2019) pmid: 31575541
- 147. Chan et al., 2017; AACR Abstract CT106/24
- 148. Kaibori M, et al. Oncotarget (2016) pmid: 27384874 149. Dumbrava EI, et al. JCO Precis Oncol (2018) pmid:
- 31123723 150. Packer LM, et al. Cancer Discov (2015) pmid: 25847957
- 151. Cerami E. et al. Cancer Discov (2012) pmid: 22588877
- 152. Gao J, et al. Sci Signal (2013) pmid: 23550210
- 153. Desnoyers LR, et al. Oncogene (2008) pmid: 17599042 154. Sawey ET, et al. Cancer Cell (2011) pmid: 21397858
- 155. Miura S. et al. BMC Cancer (2012) pmid: 22309595 156. Feng S, et al. Cancer Res. (2013) pmid: 23440425
- 157. Nagamatsu H. et al. Prostate (2015) pmid: 25854696 158. Xie MH, et al. Cytokine (1999) pmid: 10525310

APPENDIX

References

- 159. Int. J. Oncol. (2002) pmid: 12429977
- 160. Kan Z, et al. Genome Res. (2013) pmid: 23788652
- 161. Dumbrava et al., 2018; doi/full/10.1200/PO.18.00100
- 162. Tekin M, et al. Am. J. Hum. Genet. (2007) pmid:
- 163. Arao T, et al. Hepatology (2013) pmid: 22890726
- 164. Yamada T, et al. BMC Cancer (2015) pmid: 25885470
- 165. Kratochwil K. et al. Genes Dev. (2002) pmid: 12502739
- 166. Scherz PJ, et al. Science (2004) pmid: 15256670
- 167. Zaharieva BM, et al. J. Pathol. (2003) pmid: 14648664
- 168. Arai H, et al. Cancer Genet. Cytogenet. (2003) pmid: 14499691
- 169. Ribeiro IP, et al. Tumour Biol. (2014) pmid: 24477574
- 170. Schulze K, et al. Nat. Genet. (2015) pmid: 25822088
- 171. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
- 172. Robinson G, et al. Nature (2012) pmid: 22722829
- 173. Ho AS, et al. Nat. Genet. (2013) pmid: 23685749
- 174. Grasso CS, et al. Nature (2012) pmid: 22722839
- 175. Van der Meulen J, et al. Blood (2015) pmid: 25320243
- 176. Wang L, et al. Nat Commun (2013) pmid: 23792809
- 177. Kim JH, et al. Cancer Res. (2014) pmid: 24491801
- 178. Shen Y, et al. BMC Cancer (2012) pmid: 23057811
- 179. van Haaften G, et al. Nat. Genet. (2009) pmid:
- 19330029 180. Kumar A, et al. Nat. Med. (2016) pmid: 26928463
- 181. Bivona TG, et al. Nature (2011) pmid: 21430781
- 182. Giannikopoulos et al., 2014; ASCO Abstract 8083
- 183. Zhao Z, et al. Int. J. Mol. Med. (2014) pmid: 25215581
- 184. Geng P, et al. Medicine (Baltimore) (2015) pmid:
- 185. Zhang M, et al. Med. Sci. Monit. (2015) pmid: 26488500
- 186. Weniger MA, et al. Semin. Cancer Biol. (2016) pmid: 27221964
- 187. Liu X, et al. Cancer Genet. Cytogenet. (2010) pmid: 20193848
- 188. Lake A. et al. Int. J. Cancer (2009) pmid: 19507254
- 189. Birnstiel ML, et al. Arch Biol (Liege) (1965) pmid: 5858823
- 190. Cabannes E, et al. Oncogene (1999) pmid: 10340377
- 191. Bredel M. et al. N. Engl. J. Med. (2011) pmid: 21175304 192. Patanè M, et al. Mol. Cancer (2013) pmid: 24330732
- 193. Zack Tl. et al. Nat. Genet. (2013) pmid: 24071852
- 194. Beroukhim R, et al. Nature (2010) pmid: 20164920
- 195. Hsu DS, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid:
- Yang L, et al. J Zhejiang Univ Sci B (2012) pmid: 196. 23125078
- 197. Landa I, et al. J. Clin. Invest. (2016) pmid: 26878173
- 198. Jiao Y, et al. J. Pathol. (2014) pmid: 24293293
- 199. Jones S, et al. Nat Commun (2014) pmid: 25233892 200. Nakamura N, et al. Mod. Pathol. (2002) pmid: 12379752
- 201. Moldvay J, et al. Pathol. Oncol. Res. (2004) pmid: 15188024
- Gilbert-Sirieix M, et al. PLoS ONE (2011) pmid: 21814573
- Robens J, et al. Am. J. Surg. Pathol. (2010) pmid: 203.
- 204. Ni YB, et al. Histopathology (2014) pmid: 24111789
- 205. Tsai LH, et al. Oncogene (2014) pmid: 23995788
- 206. Tan D, et al. Hum. Pathol. (2003) pmid: 12827614
- 207. Haque AK, et al. Appl. Immunohistochem. Mol. Morphol. (2002) pmid: 12051626 208. Pelosi G, et al. Am. J. Surg. Pathol. (2001) pmid:
- 11224607
- 209. Yamaguchi T, et al. Cancer Cell (2013) pmid: 23763999 210. J. Biol. Chem. (2013) pmid: 23818522
- 211. Hamdan H, et al. Biochim. Biophys. Acta (1998) pmid:

- 212. Kwei KA, et al. Oncogene (2008) pmid: 18212743
- 213. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315 214. Bridges KA, et al. Clin. Cancer Res. (2011) pmid:
- 21799033 215. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid:
- 21389100
- 216. Osman AA, et al. Mol. Cancer Ther. (2015) pmid: 25504633
- 217. Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
- 218. Xu L, et al. Mol. Med. (2001) pmid: 11713371
- 219. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
- 220. Kim SS, et al. Nanomedicine (2015) pmid: 25240597
- 221. Pirollo KF, et al. Mol. Ther. (2016) pmid: 27357628
- 222. Haidenberg et al., 2012; ASCO Abstract e15010
- 223. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
- 224. Moore et al., 2019: ASCO Abstract 5513
- 225. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27998224
- 226. Oza et al., 2015; ASCO Abstract 5506
- 227. Lee J, et al. Cancer Discov (2019) pmid: 31315834
- 228. Méndez E, et al. Clin. Cancer Res. (2018) pmid:
- 229. Ma CX, et al. J. Clin. Invest. (2012) pmid: 22446188
- 230. Lehmann S, et al. J. Clin. Oncol. (2012) pmid: 22965953
- 231. Mohell N, et al. Cell Death Dis (2015) pmid: 26086967
- 232. Fransson Å, et al. J Ovarian Res (2016) pmid: 27179933
- 233. Gourley et al., 2016; ASCO Abstract 5571
- 234. Kwok M, et al. Blood (2016) pmid: 26563132
- 235. Boudny M, et al. Haematologica (2019) pmid: 30975914
- 236. Dillon MT, et al. Mol. Cancer Ther. (2017) pmid: 28062704
- 237. Middleton FK, et al. Cancers (Basel) (2018) pmid: 30127241
- 238. Mogi A, et al. J. Biomed. Biotechnol. (2011) pmid:
- 239. Tekpli X, et al. Int. J. Cancer (2013) pmid: 23011884
- 240. Vignot S, et al. J. Clin. Oncol. (2013) pmid: 23630207
- 241. Maeng CH, et al. Anticancer Res. (2013) pmid: 24222160
- 242. Cortot AB, et al. Clin Lung Cancer (2014) pmid: 24169260
- 243. Itakura M, et al. Br. J. Cancer (2013) pmid: 23922113
- 244. Dong ZY, et al. Clin. Cancer Res. (2017) pmid: 28039262
- 245. Seo JS, et al. Genome Res. (2012) pmid: 22975805
- 246. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675 Joerger AC, et al. Annu. Rev. Biochem. (2008) pmid: 18410249
- 248. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12826609
- 249. Kamada R, et al. J. Biol. Chem. (2011) pmid: 20978130
- 250. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) pmid: 28472496
- 251. Yamada H, et al. Carcinogenesis (2007) pmid: 17690113
- 252. Landrum MJ, et al. Nucleic Acids Res. (2018) pmid:
- 253. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
- 254. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
- 255. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
- 256. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
- 257. Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
- 258. Lalloo F. et al. Lancet (2003) pmid: 12672316
- 259. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713
- 260. Jaiswal S. et al. N. Engl. J. Med. (2014) pmid: 25426837
- 261. Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838

- 262. Xie M. et al. Nat. Med. (2014) pmid: 25326804
- 263. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
- 264. Severson EA, et al. Blood (2018) pmid: 29678827
- 265. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
- 266. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
- 267. Chabon JJ, et al. Nature (2020) pmid: 32269342
- 268. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
- 269. Wu YL, et al. Lancet Oncol. (2014) pmid: 24439929
- 270. Passaro et al., 2019; ELCC Abstract 1150
- 271. Audet et al., 2013; ASCO Abstract 6041 272. Lau SC, et al. Clin Lung Cancer (2019) pmid: 31178389
- 273. Paz-Ares L, et al. Ann. Oncol. (2017) pmid: 28426106
- 274. Thongprasert S, et al. Lung Cancer Manag (2019) pmid: 31807143
- 275. Januszewski et al., 2018; IASLC WCLC Abstract P1.13-17
- 276. Suzuki et al., 2018; IASLC WCLC Abstract P1.01-92
- 277. Chang et al., 2018; IASLC WCLC Abstract P1.01-11
- 278. Llinás-Quintero N, et al. Case Rep Oncol Med (2019) pmid: 31637072
- 279. Miller VA, et al. Lancet Oncol. (2012) pmid: 22452896
- 280. Chen X, et al. Lung Cancer (2013) pmid: 23664448
- 281. Katakami N, et al. J. Clin. Oncol. (2013) pmid: 23816963
- 282. Landi L, et al. Clin Lung Cancer (2014) pmid: 25242668
- 283. De Grève J, et al. Lung Cancer (2015) pmid: 25682316
- 284. Yang JC, et al. Lancet Oncol. (2015) pmid: 26051236
- 285. Horn L, et al. Lung Cancer (2017) pmid: 29110849 286. Yamamoto N, et al. Adv Ther (2020) pmid: 31863283
- 287. Soria JC, et al. Lancet Oncol. (2015) pmid: 26156651
- 288. Dziadziuszko R, et al. J Thorac Oncol (2019) pmid: 30825613
- 289. Lai WV, et al. Eur. J. Cancer (2019) pmid: 30685684 290. Greulich H, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) pmid: 22908275
- 291. Gow CH, et al. J Thorac Oncol (2015) pmid: 26134234
- 292. Mazières J, et al. Ann. Oncol. (2016) pmid: 26598547
- 293. Mazières J, et al. J. Clin. Oncol. (2013) pmid: 23610105 294. De Grève J, et al. Lung Cancer (2012) pmid: 22325357
- 295. Li BT, et al. Lung Cancer (2015) pmid: 26559459
- 296. Costa DB, et al. J Thorac Oncol (2016) pmid: 26964772
- 297. Yuan B. et al. Front Oncol (2020) pmid: 32477948
- 298. Fang W, et al. Oncologist (2019) pmid: 31748336
- 299. Schuler M, et al. Ann. Oncol. (2016) pmid: 26646759 300. Opsomer RJ, et al. Acta Urol Belg (1985) pmid: 2986437
- 301. Wu et al., 2018; WCLC abstract MA26.11
- 302. Ramalingam SS, et al. Ann. Oncol. (2016) pmid:
- 26768165 303. Yu HA, et al. Lung Cancer (2017) pmid: 29191595
- 304. Reckamp KL, et al. Cancer (2014) pmid: 24501009
- 305. Jänne PA, et al. Clin. Cancer Res. (2011) pmid: 21220471 306. van Geel RMJM, et al. Br. J. Cancer (2020) pmid:
- 32147669
- **307.** Jänne PA, et al. J Thorac Oncol (2016) pmid: 26899759
- 308. Cappuzzo F, et al. Lancet Oncol. (2010) pmid: 20493771
- 309. Zhong WZ, et al. J. Clin. Oncol. (2019) pmid: 31194613 310. Petrelli F, et al. Clin Lung Cancer (2012) pmid:
- 22056888
- 311. Yang JJ, et al. Br. J. Cancer (2017) pmid: 28103612 312. Lee CK, et al. J. Natl. Cancer Inst. (2017) pmid: 28376144
- 313. Nakagawa K, et al. Lancet Oncol. (2019) pmid: 31591063 314. Stinchcombe TE, et al. JAMA Oncol (2019) pmid:
- 31393548
- 315. Truini A, et al. Clin. Cancer Res. (2019) pmid: 31182434 316. Shepherd FA, et al. N. Engl. J. Med. (2005) pmid:

APPENDIX

References

- 317. Han JY, et al. J. Clin. Oncol. (2012) pmid: 22370314
- **318.** Maemondo M, et al. N. Engl. J. Med. (2010) pmid: 20573926
- **319.** Mitsudomi T, et al. Lancet Oncol. (2010) pmid: 20022809
- **320.** Mok TS, et al. N. Engl. J. Med. (2009) pmid: 19692680
- 321. Qi WX, et al. Curr Med Res Opin (2015) pmid: 25329826322. Zhao H, et al. J Thorac Oncol (2015) pmid: 25546556
- 323. Wang J, et al. Int. J. Cancer (2019) pmid: 30255937
- **324.** Baik CS, et al. J Thorac Oncol (2015) pmid: 26398831
- **325.** Yoshioka H, et al. Ann. Oncol. (2019) pmid: 31553438
- **326.** Fukuoka M, et al. J. Clin. Oncol. (2011) pmid: 21670455
- 327. Noronha V, et al. J. Clin. Oncol. (2019) pmid: 31411950
- **328.** Hosomi Y, et al. J. Clin. Oncol. (2020) pmid: 31682542 **329.** Sutiman N, et al. J Thorac Oncol (2017) pmid: 27908825
- **330.** Gibbons DL, et al. J Thorac Oncol (2016) pmid: 27198414
- 331. Alanazi A, et al. Lung Cancer Manag (2020) pmid:

33318755

- 332. Kim et al., 2021; DOI: 10.1200/PQ.20.00296
- **333.** Ramalingam SS, et al. N. Engl. J. Med. (2019) pmid: 31751012
- **334.** Herbst et al., 2020; ASCO Abstract LBA5
- 335. Yu HA, et al. JAMA Oncol (2020) pmid: 32463456
- **336.** Oxnard GR, et al. Ann. Oncol. (2020) pmid: 32139298

